

## Comparative of some growth, biochemical, and hormonal responses of galled branches of two species of willow trees

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### Abstract

The witches' broom of the willow trees is induced by photoplasma in the galled branches. The aim of the study is to evaluate the changes in growth and physiological factors and the plant growth promoting hormones (Auxin, Gibberellin, and Cytokinin) in leaf witches' broom branches compared with those healthy symptomless branches. The samples were collected from the leaves of ungalled and galled branches of both willow trees in Najvan Park, Isfahan. Results showed that growth parameters, total chlorophyll, and total soluble carbohydrates were decreased in the leaves of witches' broom branches in both species. However, the relative water content and polyphenol oxidase activity did not show a significant change as compared with those of ungalled branches. In contrast, the ascorbate peroxidase activity did not show a significant difference between the leaves of ungalled and witches' broom branches in *S. babylonica*, but its activity was decreased in witches' broom branches of *S. alba*. The cytokinin content was increased (more than 50%) in the leaves of witches' broom branches of both species, but the auxin and GA contents in the leaves of witches' broom branches of *S. alba* and *S. babylonica* were significantly decreased (30.6% and 33.6% respectively) as compared with those of healthy control leaves. The concentration of total chlorophyll and water-soluble sugar decreased due to oxidative stress induced by phytoplasma. Reduced growth with no significant change in the relative water content of galled branches in both willow trees is accompanied by the specific hormonal changes of each species which is formed of specific galls for each species.

### Introduction

The genus Willow (*Salix*) are shrubs and trees that usually grow in most parts of the earth. Plant galls are neoformed organs that develop under the influence of galling organisms. Insects, nematodes, viruses, bacteria, mycoplasma, fungi, other plants, and mites are typically considered gall inducers. They cause structural and metabolic changes in their host plants, leading to the formation of the gall organ. Three hypotheses nutrition, microenvironment, and enemy have been proposed for forming galls by

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insects. The gall formation depends on the level of gene transcription interactions that exist between gall inducers and plants. The function of these genes causes changes in growth and development processes, cytoskeleton organization, cell walls, metabolic pathways, and plant hormone pathways. Recently, *Candidatus phytoplasma trifolii* caused willow catkin gall on *Salix babylonica* and *S. alba* trees, characterized by a unique case of neoplastic formation like witches' broom structure. The aim of the study is to evaluate the changes in growth and physiological factors and the plant growth-promoting hormones (Auxin, Gibberellin, and Cytokinin) in leaf witches' broom branches compared with those healthy symptomless branches.

#### Material and methods

Leaf samples of trees with gall and their symptomless adjacent branches were collected from Najvan Park, Isfahan province, Iran. The length and width of the leaves were measured and recorded. Fresh and dry weights and relative water content (RWC) of leaves were measured. Total chlorophyll was measured according to the Arnon assay. Water soluble carbohydrates were assessed using the method of Dubois (1956). Activities of ascorbate peroxidase (APX) and polyphenol oxidase (PPO) were measured by the methods of Asada and Takahashi (1987) and Kar & Mishra (1976), respectively. High-performance liquid chromatography (HPLC) was used to analyze plant growth promoting hormones.

#### Results and discussion

The gall formation in *Salix* species is caused by *Candidatus phytoplasma trifolii*. The gall formation was accompanied by a reduction in the growth of leaves and shoots, which was represented by smaller leaves and shorter internodes. The gallers used nutritional resources of gall and adverse effects on the growth and survival of the host plants. Results showed that relative water content (RWC) showed no significant changes between the witches' broom structure compared with those of healthy control leaves in both *Salix* species. Reduction in growth without change in RWC is due to the modification in the expression of genes which is indicated by smaller leaves in gall. Total chlorophyll was reduced in all galled tissue of both *Salix* species. The reactive oxygen species (ROS) generated to respond to the biological stress in chloroplast can induce oxidative damage to the photosynthetic pigments which leads to the reduction of soluble sugars in the galled leaves. Polyphenol oxidase (PPO) activity did not show a significant change as compared with those of healthy branches in both galled leaves *Salix* species. In contrast, the ascorbate peroxidase activity did not show a significant difference between the leaves of galled and healthy branches in *S. babylonica*. However, its activity was decreased in galled branches of *S. alba*, which indicated the activity of these enzymes is not a priority in both galled leaves *Salix* species. The cytokinin content was increased in the leaves of witches' broom branches of both species. However, the auxin and GA contents in the leaves of witches' broom branches of *S. alba* and *S. babylonica* were significantly decreased by 30.6% and 33.6% respectively as compared with those of healthy control leaves. Elevated cytokinin content in both willow species can be introduced as the main factor in increasing the divisions of witches' broom branches which is coordinated with the particular hormonal changes each species which generates special galls them.

### Conclusions

*Candidatus* phytoplasma trifolii reduced the growth of infected tissues characterized by smaller leaves and shorter internodes, followed by the formation of gall structure in both *Salix* species. According to the results, the high accumulation of ROS in response to the gall formation was accompanied by a reduction of total chlorophyll and water soluble carbohydrates. Altogether, it seems that the adverse effects of gallers on *Salix* species refer partly to oxidative stress, which is synchronized with the specific hormonal changes of each species. These events cause the creation of special galls for each species.

Keywords: Enzymes activity, Gall, Growth promoting hormones, Total chlorophyll, Willow



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Figure 1- Picture of willow trees, Salix alba (right side) and Salix babylonica (left side) which one of their galls branches is shown at the bottom of the pictures (C, D)

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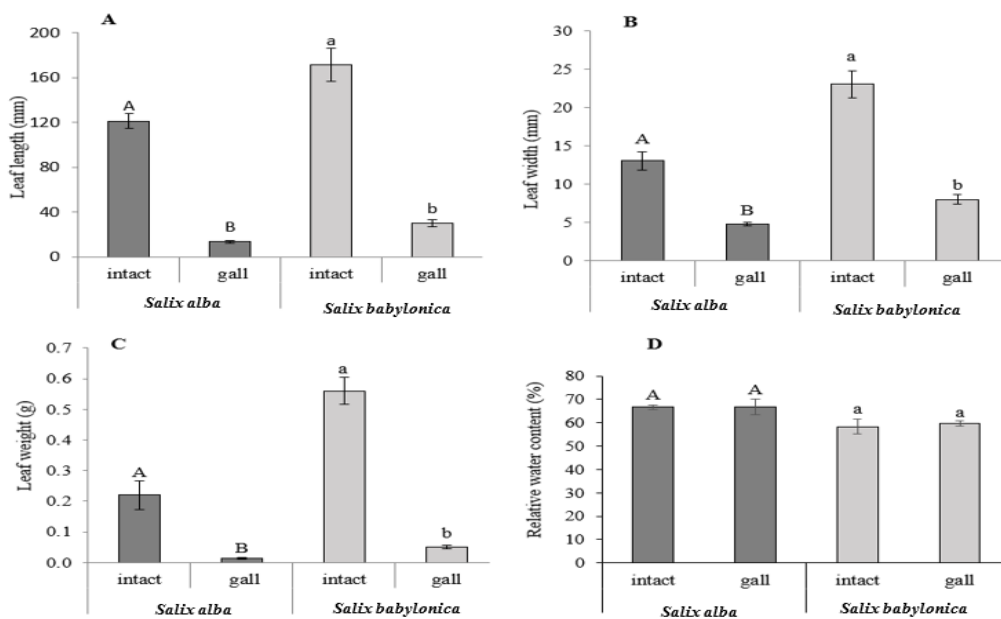
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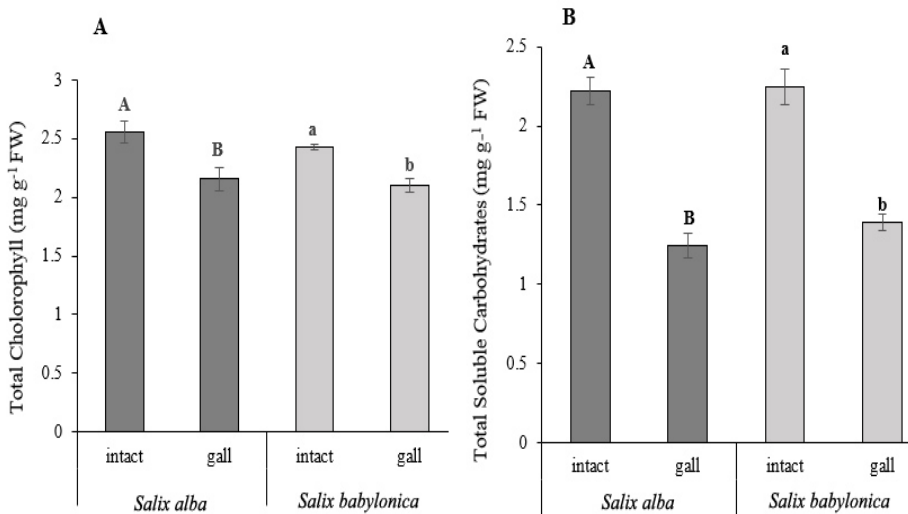


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Figure 2- Comparison of average length(A), width (B), weight (C) and relative water content of leaves (D) of ungalled and galled branches on two species of *Salix*. Values are expressed by the mean ± standard error. In each species, different letters (uppercase for *S. alba* and lowercase for *S. babylonica*) indicate statistically significant differences in t-test (P < 0.05).



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Figure 3- Comparison of average total chlorophyll (A) and total soluble carbohydrates (B) of ungalled and galled branches on two species of *Salix*. Values are expressed by the mean ± standard error. In each species, different letters (lowercase for *S. alba* and uppercase for *S. babylonica*) indicate statistically significant differences in t-test ( $P < 0.05$ ).

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Figure 4- Comparison of the average activity of enzymes ascorbate oxidase (APX) (A) and polyphenol oxidase (PPO) activity of leaves (B) of ungalled and galled branches on two species of *Salix* values are expressed by the mean  $\pm$  standard error. In each species, different letters (lowercase for *Salix alba* and uppercase for *Salix babylonica*) indicate statistically significant differences in t-test ( $P < 0.05$ ).

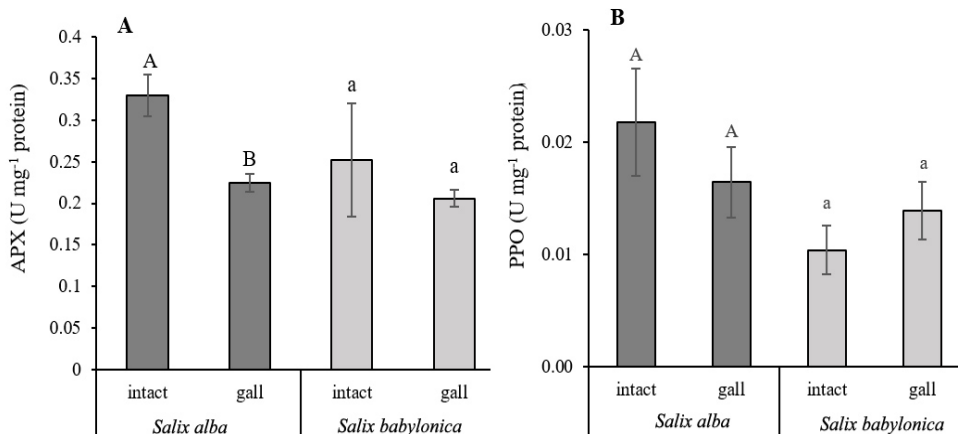


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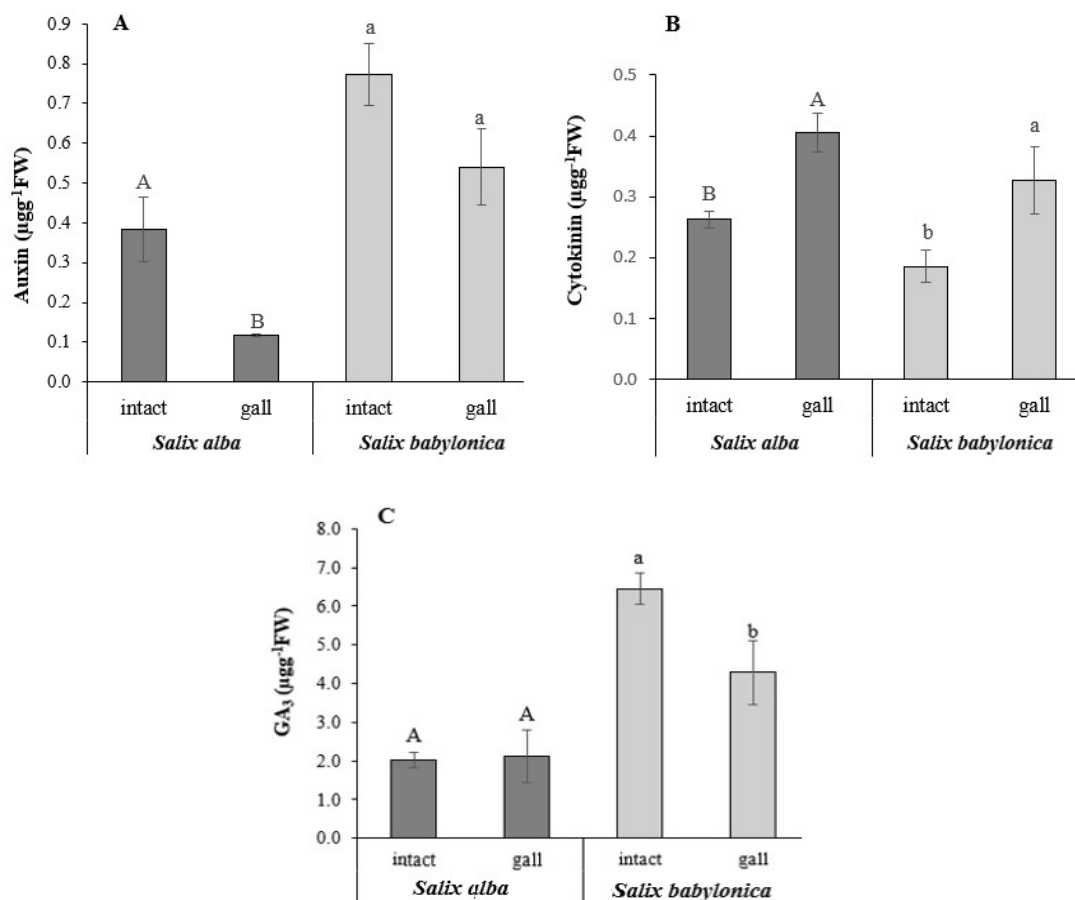


Figure 5- Comparison of average the hormones auxin (A), cytokinin (B) and gibberellin (GA) of ungalged and galged branches on two species of Salix. Values are expressed by the mean ± standard error. In each species, different letters (lowercase for *S. alba* and uppercase for *S. babylonica*) indicate statistically significant differences in t-test ( $P < 0.05$ ).

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- multinucleate and hypertrophied feeding cells. *Journal of Insect Physiology*, 84, 60-69. <https://doi.org/10.1016/j.jinsphys.2015.07.013>
- Gailite, A., Andersone, U. & Ievinsh, G. (2005). Arthropod-induced neoplastic formations on trees change photosynthetic pigment levels and oxidative enzyme activities. *Journal of Plant Interactions*, 1(1), 61-67. <https://doi.org/10.1080/17429140500254728>
- Ghayeb Zamharir, M. (2018). Association of 'Candidatus Phytoplasma trifolii'-related strain with white willow proliferation in Iran. *Australasian Plant Disease Notes*, 13(1), 1-4. <https://doi.org/10.1007/s13314-018-0300-y>
- Gheysen, G. & Mitchum, M. G. (2019). Phytoparasitic nematode control of plant hormone pathways. *Plant Physiology*, 179(4), 1212-1226. <https://doi.org/10.1104/pp.18.01067>
- Giron, D., Huguet, E., Stone, G. N. & Body, M. (2016). Insect-induced effects on plants and possible effectors used by gall-forming insects to manipulate their host plant. *Journal of Insect Physiology*, 84, 70-89. <https://doi.org/10.1016/j.jinsphys.2015.12.009>
- Guedes, L. M., Sanhueza, C., Torres, S., Figueroa, C., Gavilán, E., Pérez, C. I. & Aguilera, N. (2023). Gall-inducing *Eriophyes tiliae* stimulates the metabolism of *Tilia platyphyllos* leaves towards oxidative protection. *Plant Physiology and Biochemistry*, 192, 25-36. <https://doi.org/10.1016/j.plaphy.2022.12.014>
- Guiget, A., Takeda, S., Hirano, T., Issa, & Sato, M. H. (2021). Recent Progress Regarding the Evolution and Molecular Aspect of Insect Gall Formation. <https://doi.org/10.20944/preprints202106.0494.v1>
- (2022). Antioxidant capacity of *Salix alba* (Fam. Salicaceae) and influence of heavy metal accumulation. *Horticulturae*, 8(7), 642. <https://doi.org/10.3390/horticulturae8070642>
- Barna, B. & Pogány, M. (2001). Antioxidant enzymes and membrane lipid composition of disease resistant tomato plants regenerated from crown galls. *Acta Physiologiae Plantarum*, 23, 273-277. <https://doi.org/10.1007/s11738-0033x>
- Bohlmann, H. & Sobczak, M. (2014). The plant cell wall in the feeding sites of cyst nematodes. *Frontiers in Plant Science*, 5, 89. <https://doi.org/10.3389/fpls.2014.00089>
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(2), 248-254. [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- Cambier, S., Ginis, O., Moreau, S. J., Gayral, P., Heman, J., Stone, G. N., & Drezen, J. M. (2019). Gall wasp transcriptomes unravel potential effectors involved in molecular dialogues with oak and rose. *Frontiers in Plant Physiology*, 10, 926. <https://doi.org/10.3389/fphys.2019.00926>
- Favery, B., Dubreuil, G., Chen, M. S., Giron, D. & Abad, P. (2020). Gall-inducing parasites: convergent and conserved strategies of plant manipulation by insects and nematodes. *Annual Review of Phytopathology*, 58, 1-22. <https://doi.org/10.1146/annurev-phyto-010820012722>
- Favery, B., Quentin, M., Jaubert, S. & Abad, P. (2016). Gall-forming root-knot nematodes hijack key plant cellular functions to induce

- activities during rice leaf senescence. *Plant Physiology*, 52(2), 315-319. <https://doi.org/10.1104/pp.57.2.315>
- Kerpen, L., Niccolini, L., Licausi, F., Van Dongen, J. T. & Weits, D. A. (2019). Hypoxic conditions in crown galls induce plant anaerobic responses that support tumor proliferation. *Frontiers in Plant Science*, 10, 56. <https://doi.org/10.3389/fpls.2019.00056>
- Khadhair, A. H. & Hiruki, C. (1995). The molecular genetic relatedness of willow witches'-broom phytoplasma to the clover proliferation group. *Proceedings of the Japan Academy, Series B*, 71(1), 145-147. <https://doi.org/10.2183/pjab.71.145>
- Khajuria, C., Wang, H., Liu, X., Wheeler, S., Reese, J. C., El Bouhssini, M., & Chen, M. S. (2013). Mobilization of lipids and fortification of cell wall and cuticle are important in host defense against Hessian fly. *BMC Genomics*, 14(1), 1-16. <https://doi.org/10.1186/1472-1164-14-423>
- Kmieć, K., Kot, I., Rubinowska, K., Górská-Drabik, E., Golan, K. & Sytykiewicz, H. (2022). The variation of selected physiological parameters in elm leaves (*Ulmus glabra* Huds.) infested by gall inducing aphid. *Plants*, 11(3), 244. <https://doi.org/10.3390/plants11030244>
- Kot, I., Sempruch, C., Rubinowska, K. & Michałek, W. (2020). Effect of *Neuroterus quercusbaccarum* galls on physiological and biochemical response of *Quercus robur* leaves. *Bulletin of Entomological Research*, 110(1), 34-43. <https://doi.org/10.1017/S0007485319000221>
- Kreslavski, V., Ivanov, A., Shmarev, A., Hajhashemi, S. & Ehsanpour, A. (2014). Antioxidant response of *Stevia rebaudiana* to polyethylene glycol and paclobutrazol treatments under in vitro culture. *Applied Biochemistry and Biotechnology*, 172(8), 4038-4052. <https://doi.org/10.1007/s12010-014-0791-8>
- Harris, M., Freeman, T., Rohfritsch, O., Anderson, K., Payne, S. & Moore, J. (2006). Virulent Hessian fly (Diptera: Cecidomyiidae) larvae induce a nutritive tissue during compatible interactions with wheat. *Annals of the Entomological Society of America*, 99(2), 305-316. [https://doi.org/10.1603/0013-8746\(2006\)099\[0305:VHFDCL\]2.0.CO;2](https://doi.org/10.1603/0013-8746(2006)099[0305:VHFDCL]2.0.CO;2)
- Huang, M. Y., Huang, W. D., Chou, H. M., Chen, C. C., Chang, Y. T. & Yang, C. M. (2014). Herbivorous insects alter the chlorophyll metabolism of galls on host plants. *Journal of Asia-Pacific Entomology*, 17(3), 431-434. <https://doi.org/10.1016/j.aspen.2014.04.004>
- Ivanova, L. A., Chetverikov, P. E., Ivanov, L. A., Tumurjav, S., Kuzmin, I. V., Desnitskiy, A. G. & Tolstikov, A. V. (2022). The effect of gall mites (Acariformes, Eriophyoidea) on leaf morphology and pigment content of deciduous trees in West Siberia. *Acarina*, 30(1), 89-98. <https://doi.org/10.21684/0132-2022-30-1-89-98>
- Ji, H., Gheysen, G., Denil, S., Lindsey, K., Topping, J. F., Nahar, K., & Van Criekinge, W. (2013). Transcriptional analysis through RNA sequencing of giant cells induced by *Meloidogyne graminicola* in rice roots. *Journal of Experimental Botany*, 64(2), 3885-3898. <https://doi.org/10.1093/jxb/ert219>
- Kar, M. & Mishra, D. (1976). Catalase, peroxidase, and polyphenoloxidase



- (2016). Manipulation of host plant cells and tissues by gall-inducing insects and adaptive strategies used by different feeding guilds. *Journal of Insect Physiology*, 84, 103-113. <https://doi.org/10.1016/j.jinsphys.2015.11.012>
- Patankar, R., Starr, G., Mortazavi, B., Oberbauer, S. F. & Rosenblum, A. (2013). The effects of mite galling on the ecophysiology of two arctic willows. *Arctic, Antarctic and Alpine Research*, 45(1), 99-106. <https://doi.org/10.1657/1934-246-45.1.99>
- Salehi-Eskandari, B., Ghaderian, S. M. & Schat, H. (2017). The role of nickel (Ni) and drought in serpentine adaptation: contrasting effects of Ni on osmoprotectants and oxidative stress markers in the serpentine endemic, *Cleome heratensis*, and the related serpentinophyte, *Cleome foliolosa*. *Plant and Soil*, 417, 183-195. <https://doi.org/10.1007/s11104-017-32509>
- Salehi-Eskandari, B. & Kaviani, M. (2015). Comparison of physiological and biochemical changes in healthy trees and willow branches gall (*Salix babylonica*). *Journal of Plant Research (Iranian Journal of Biology)*, 23(4), 885-892. [In Persian]. <https://doi.org/27512>
- Salehi-Eskandari, B., Kazemi Renani, S. & Hajihashemi, S. (2024). Evaluation of physiological and morphological responses of *Salix alba* and *Salix babylonica* to witches' broom gall. *European Journal of Plant Pathology* 169, 395-408. <https://doi.org/10.1007/s10658-024-028330>
- Samsone, I., Andersone, U. & Ievinsh, G. (2011). Gall mite *Rhabdophaga rosaria* induced rosette galls on *Salix*: morphology, photochemistry of photosynthesis and defense enzyme activity. *Environmental and Experimental Biology*, 9, 296.
- Khudyakova, A. & Kosobryukhov, A. (2022). Influence of iron nanoparticles (Fe<sub>3</sub>O<sub>4</sub> and Fe<sub>2</sub>O<sub>3</sub>) on the growth, photosynthesis and antioxidant balance of wheat plants (*Triticum aestivum*). Paper presented at the Bio Web of Conferences.
- Maruta, T., Sawa, Y., Shigeoka, S. & Ishikawa, T. (2016). Diversity and evolution of ascorbate peroxidase functions in chloroplasts: more than just a classical antioxidant enzyme? *Plant and Cell Physiology*, 57(1), 1377-1386. <https://doi.org/10.1093/pcp/pcv203>
- Meikle, R. D. (1984). *Willows and poplars of Great Britain and Ireland*
- Mohajel Kazemi, E., Alinejad, F. & Mohajjel Shoja, H. (2020). Investigation of anatomical changes induced in *Fraxinus rotundifolia* leaflets by *Psyllopsis (Psyllopsis fraxini)* infection. *Iranian Journal of Plant Biology*, 12(1), 71-84. [in Persian]. <https://doi.org/10.22108/ijpb.2020.118608.1165>
- Murata, N., Takahashi, S., Nishiyama, Y. & Allakhverdiev, S. I. (2007). Photoinhibition of photosystem II under environmental stress. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1767(6), 414-421. <https://doi.org/10.1016/j.bbabi.2006.11.019>
- Nyman, T. & Julkunen-Tiitto, R. (2000). Manipulation of the phenolic chemistry of willows by gall-inducing sawflies. *Proceedings of the National Academy of Sciences*, 97(4), 13184-13187. <https://doi.org/10.1073/pnas.230294097>
- Nyman, T., Widmer, A. & Roininen, H. (2000). Evolution of gall morphology and host-plant relationships in willow feeding sawflies (Hymenoptera: Tenthredinidae). *Evolution*, 54(2), 526-533. <https://doi.org/10.1111/j.0014-3820.2000.tb0055.x>
- Oliveira, D., Isaias, R., Fernandes, G., Ferreira, B., Carneiro, R. & Fuzaro, L.

- <https://doi.org/10.1146/annurev.ento.47.091201.145247>
- Takeda, S., Yoza, M., Amano, T., Ohshima, I., Hirano, T., Sato, M. H., & Kimura, S. (2019). Comparative transcriptome analysis of galls from four different host plants suggests the molecular mechanism of gall development. *PLoS One*, 14(10), e0223686. <https://doi.org/10.1371/journal.pone.0223686>
- Tawfeek, N., Mahmoud, M. F., Hamdan, D. I., Sobeh, M., Farrag, N., Wink, M., & El-Shazly, A. M. (2021). Phytochemistry, pharmacology and medicinal uses of plants of the genus *Salix*: an updated review. *Frontiers in pharmacology*, 12, 593856. <https://doi.org/10.3389/fphar.2021.593856>
- Tooker, J. F. & Helms, A. M. (2014). Phytohormone dynamics associated with gall insects, and their potential role in the evolution of the gall-inducing habit. *Journal of Chemical Ecology*, 40, 742-753. <https://doi.org/10.1007/s10886014-0457-6>
- Schultz, J. C., Edger, P. P., Body, M. J. & Appel, H. M. (2019). A galling insect activates plant reproductive programs during gall development. *Scientific Reports*, 9(1), 1-17. <https://doi.org/10.1038/s41598-018-38475-6>
- Shahryari, F. & Allahverdipour, T. (2018). "Candidatus *Phytoplasma trifolii*" related strain affecting *Salix babylonica* in Iran. *Australasian Plant Disease Notes*, 13(1), 1-3. <https://doi.org/10.1007/s13314-018-0321-6>
- Sherin, G., Aswathi, K. R. & Puthur, J. T. (2022). Photosynthetic functions in plants subjected to stresses are positively influenced by priming. *Plant Stress*, 100079. <https://doi.org/10.1016/j.stress.2022.100079>
- Stone, G. N. & Schönrogge, K. (2003). The adaptive significance of insect gall morphology. *Trends in Ecology and Evolution*, 18(10), 512-522. <https://doi.org/10.3390/ijms22179424>
- Stone, G. N., Schönrogge, K., Atkinson, R. J., Bellido, D. & Pujadell, J. (2002). The population biology of oak gall wasps (Hymenoptera: Cynipidae). *Annual Review of Entomology*, 47, 633-668.

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